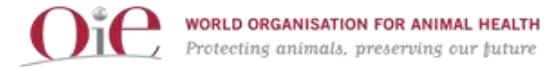




With the support of



**3rd International Symposium  
on Alternatives to Antibiotics (ATA)  
Challenges and Solutions in Animal Health and Production**

**The Berkeley Hotel, Bangkok, Thailand  
16-18 December 2019**

# **Organic Acids as Antibiotic Alternatives in Monogastric Animals**

**Dr. Chengbo Yang, Assistant Professor**

**Department of Animal Science  
Faculty of Agricultural and Food Sciences  
University of Manitoba**



# Growing demand for animal protein



7 billion people consuming on average 37 kg of meat and 83 kg of dairy per year



9 billion people consuming on average 50 kg of meat and 99 kg of dairy per year

Production increase needed by 2050:

**+ 53%**

**+ 75%**



**University  
of Manitoba**

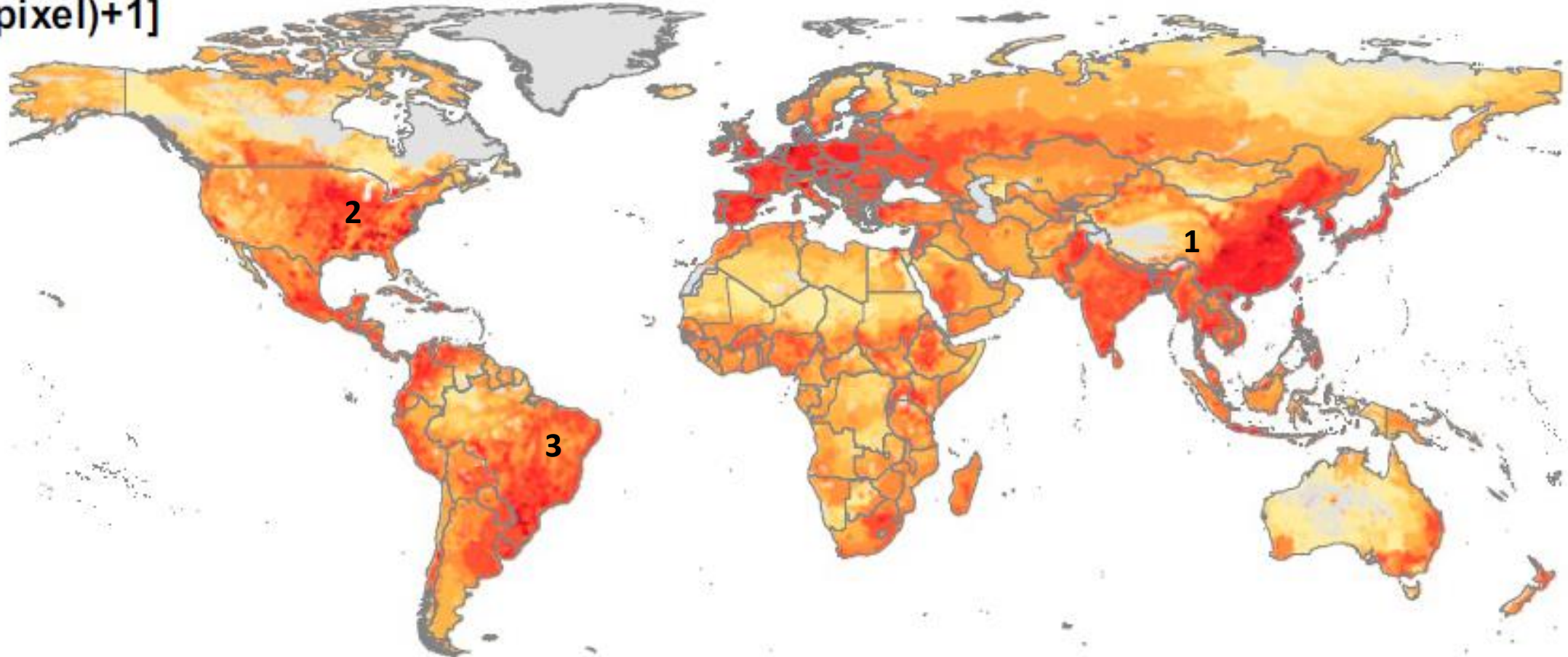
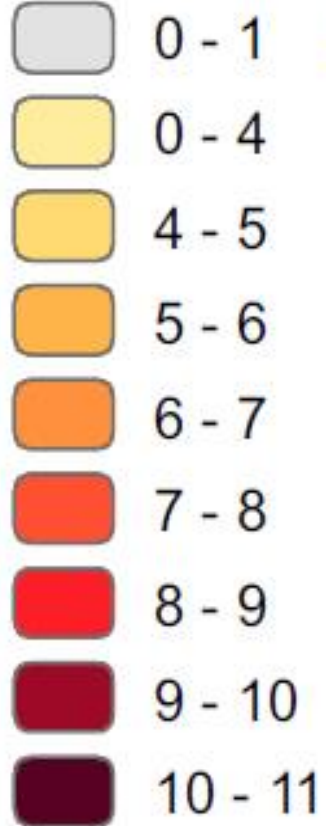
# Effect of AGP on Growth Rates in Swine (>32,000 pigs)

Class of pigs	% increase in ADG
Weanling (7-25 kg)	16.4
Growing pigs (17-49 kg)	10.6
Growing-finishing (24-84 kg)	4.2

*(Source: NRC, 1998)*

# Global trends in antimicrobial use in food animals

Log10 [(mg/pixel)+1]



1. China
2. United States
3. Brazil

63,151 tons in 2010.

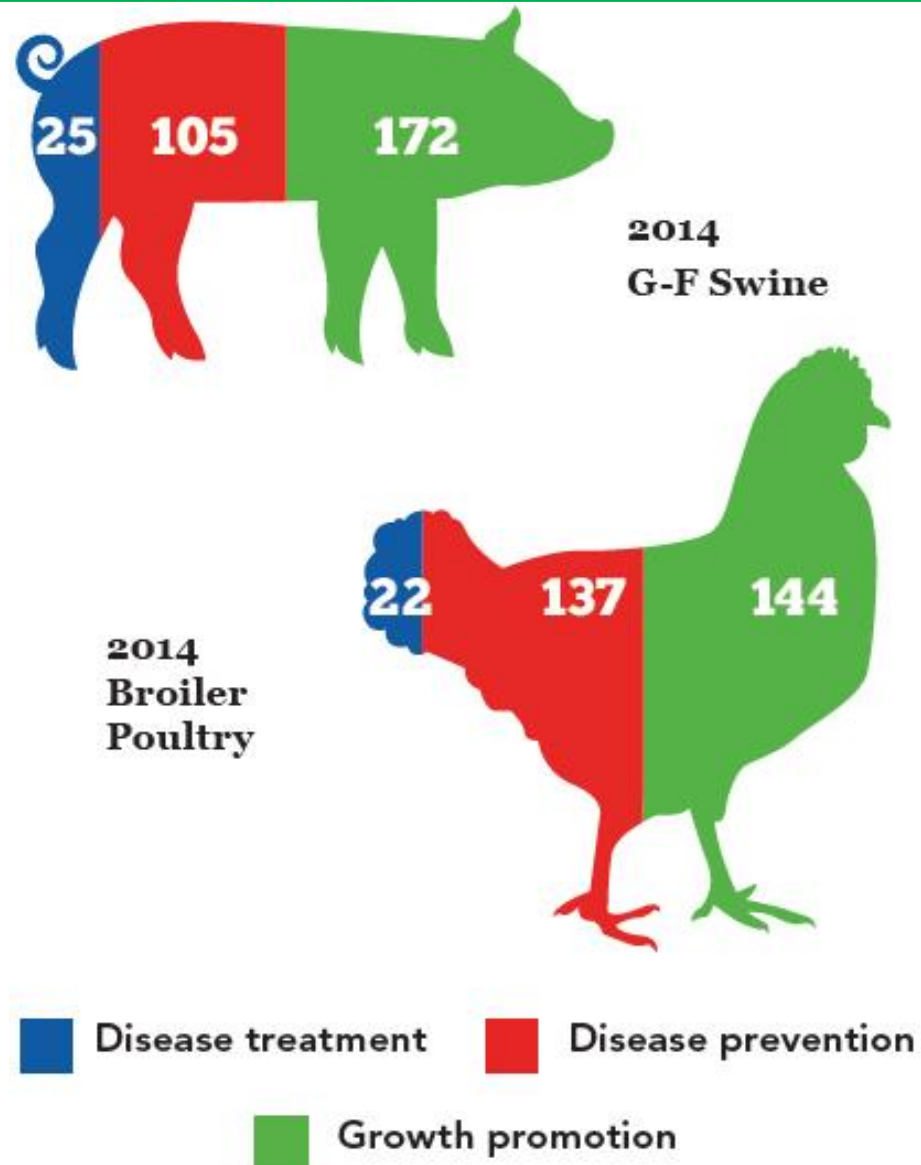
Predicted to rise dramatically by 67% by 2030.

*(Van Boeckel et al., 2015, PANS)*



**University  
of Manitoba**

# Antimicrobial use in feed adjusted for populations and weights



Source: Canadian Antimicrobial Resistance Surveillance System (CARSS) – Report 2016

# Consumers want a food system that assures food is safe, affordable, healthful and sustainable



Tyson Foods, Inc.



# The Opportunities

- ❖ **Development of better solutions to improve gut development and health in livestock; and**
- ❖ **Development of antibiotic alternatives for sustainable livestock production.**

# Alternatives to antibiotics

- ❖ Probiotics
- ❖ Prebiotics
- ❖ Synbiotics
- ❖ Feed enzymes
- ❖ Phytochemicals
- ❖ **Organic acids**
- ❖ Antimicrobial peptides
- ❖ Bacteriophage
- ❖ Bacteriophage gene products
- ❖ Antibacterial lytic enzymes
- ❖ Small interfering RNAs
- ❖ Therapeutic antibodies
- ❖ Vaccines

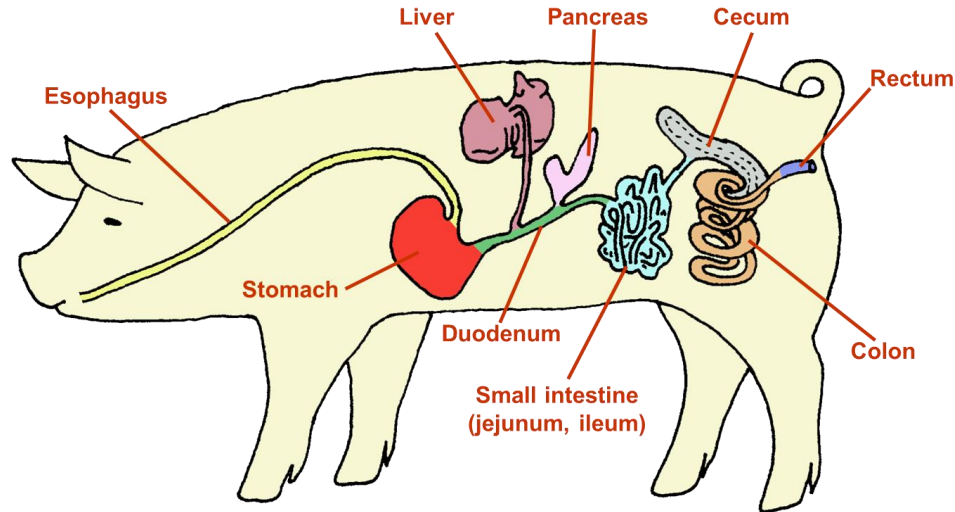




# Potential antibiotic alternatives and their estimated relative effectiveness compared to antibiotic growth promoters

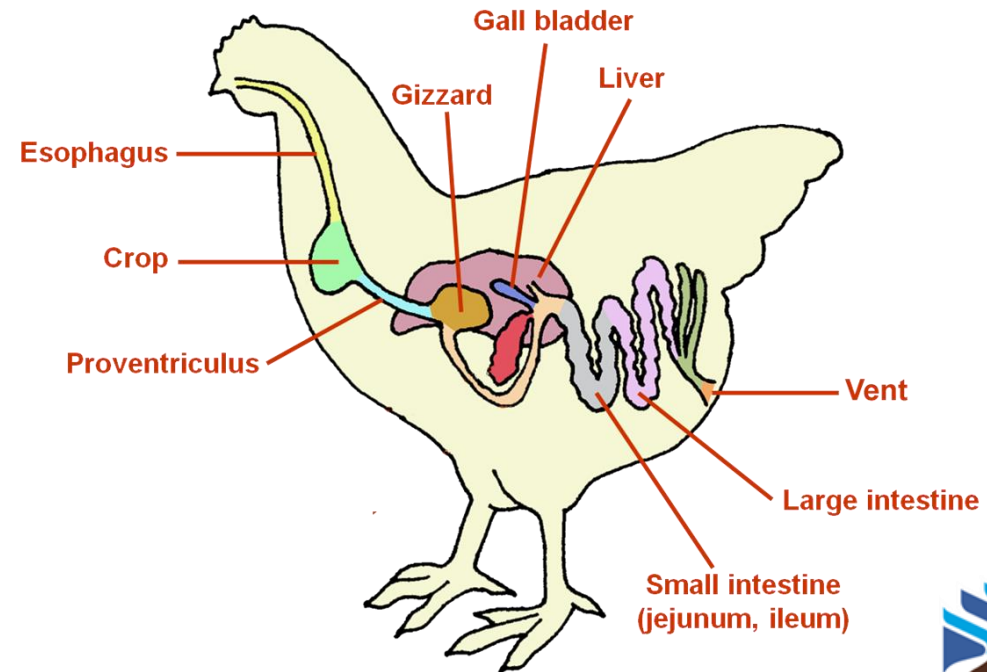
Compound	Relative Effectiveness	Comments
Antibiotic growth promotants	+++++	The standard for comparison purposes
Zinc oxide	++++	Fed at 2000 to 3000 ppm for the first two weeks post weaning. Decrease in scours and improved performance.
Copper sulfate	+++	Improved performance at 200 to 250 ppm, similar (but independent) to antibiotics. Largest effect in the nursery.
Plasma protein	+++	Increased feed intake and improved growth performance. Effects appear to be greater under unsanitary conditions.
Specific antibodies (egg yolk)	++	Limited data, but potentially promising. Results will likely depend on disease condition.
Organic acids	+++	<b>Likely most effective in newly weaned pigs. Inconsistent results. Formic acid may be most effective, but is not approved in the U.S.</b>
Direct-fed microbials	++	Suggested to promote beneficial bacteria in the gut. Inconsistent results. May depend on strain selection.
Prebiotics	++	Suggested to promote beneficial bacteria in the gut. Research with oligosaccharides have shown beneficial results.
Enzymes	++	Potential benefit through improved digestibility of feed ingredients and subsequent improved gut health.
Bioactive peptides	++	Limited research. Some peptides have antibiotic properties and could have potential benefits.
Botanicals (herbs and spices)	+	More research is necessary, There are many potential products.
Essential oils	+	More research is necessary.
Fermented liquid feeds	+	Fermentation will produce acids that can help in the maintenance of gut pH.

# Non-ruminant (monogastrics)



Segment	pH
Stomach	2.6 to 4.2
Duodenum	5 to 6
Jejunum	6.0 to 7.0
Ileum	6.0 to 7.4
Cecum/colon	6.4 to 7.5

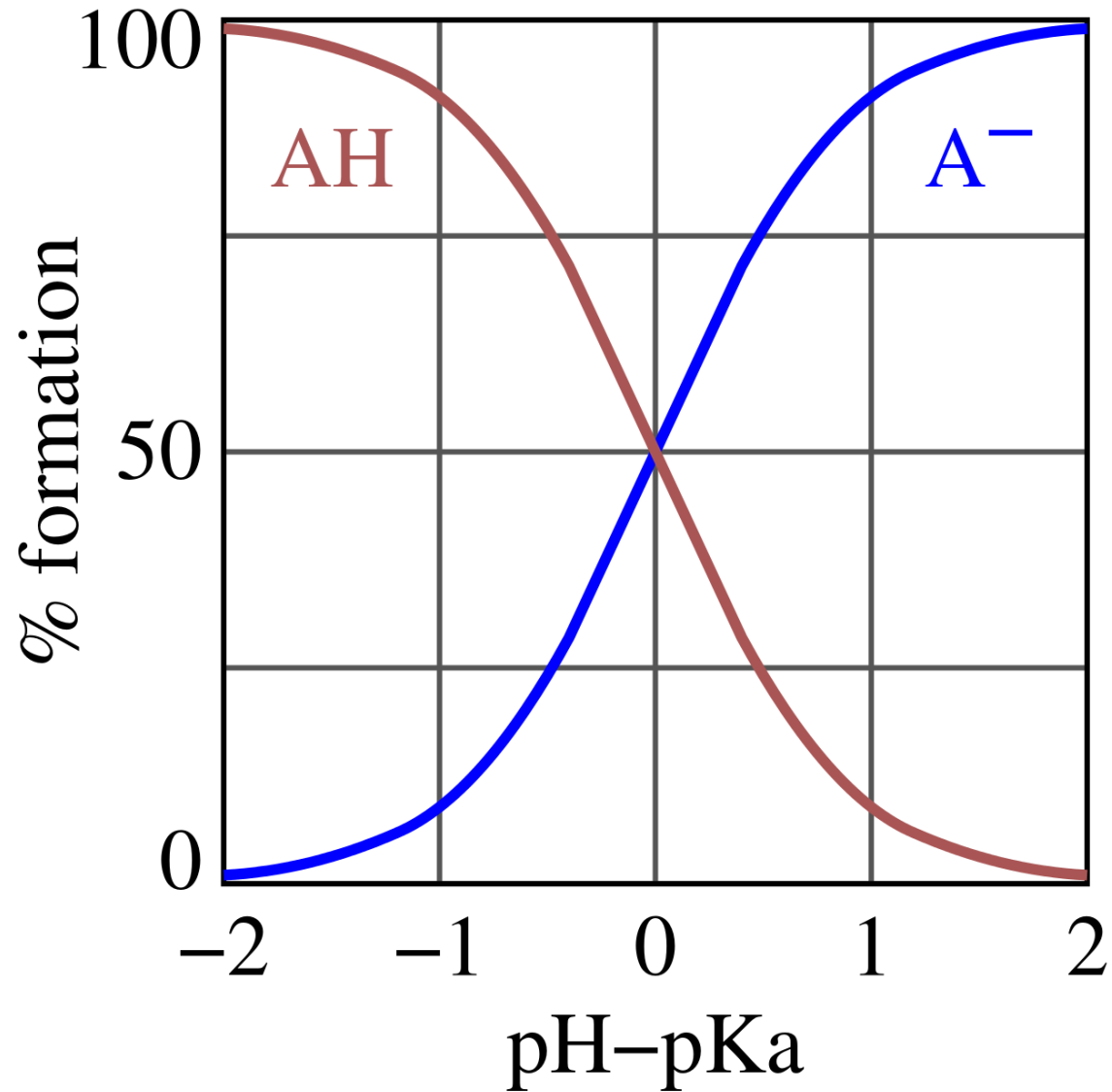
Segment	pH
Crop	5.5
Proventriculus/gizzard	2.5 to 3.5
Duodenum	5 to 6
Jejunum	6.5 to 7.0
Ileum	7.0 to 7.5
Cecum/colon	8



# Organic acids

- **Organic acids:** formic acid, citric acid, propionic acid, fumaric acid, humic acid, medium chain fatty acids (e.g. **lauric acid**).
- **Salt of organic acids:** sodium formate, sodium butyrate
- **Organic acid derivative:** Tributyrin
- **Combination of organic acids and inorganic acids** has lower inclusion levels.
- **Protected organic acids or slowly release organic acids** may have a better performance with lower inclusion levels.

# Association and dissociation equilibrium of organic acids



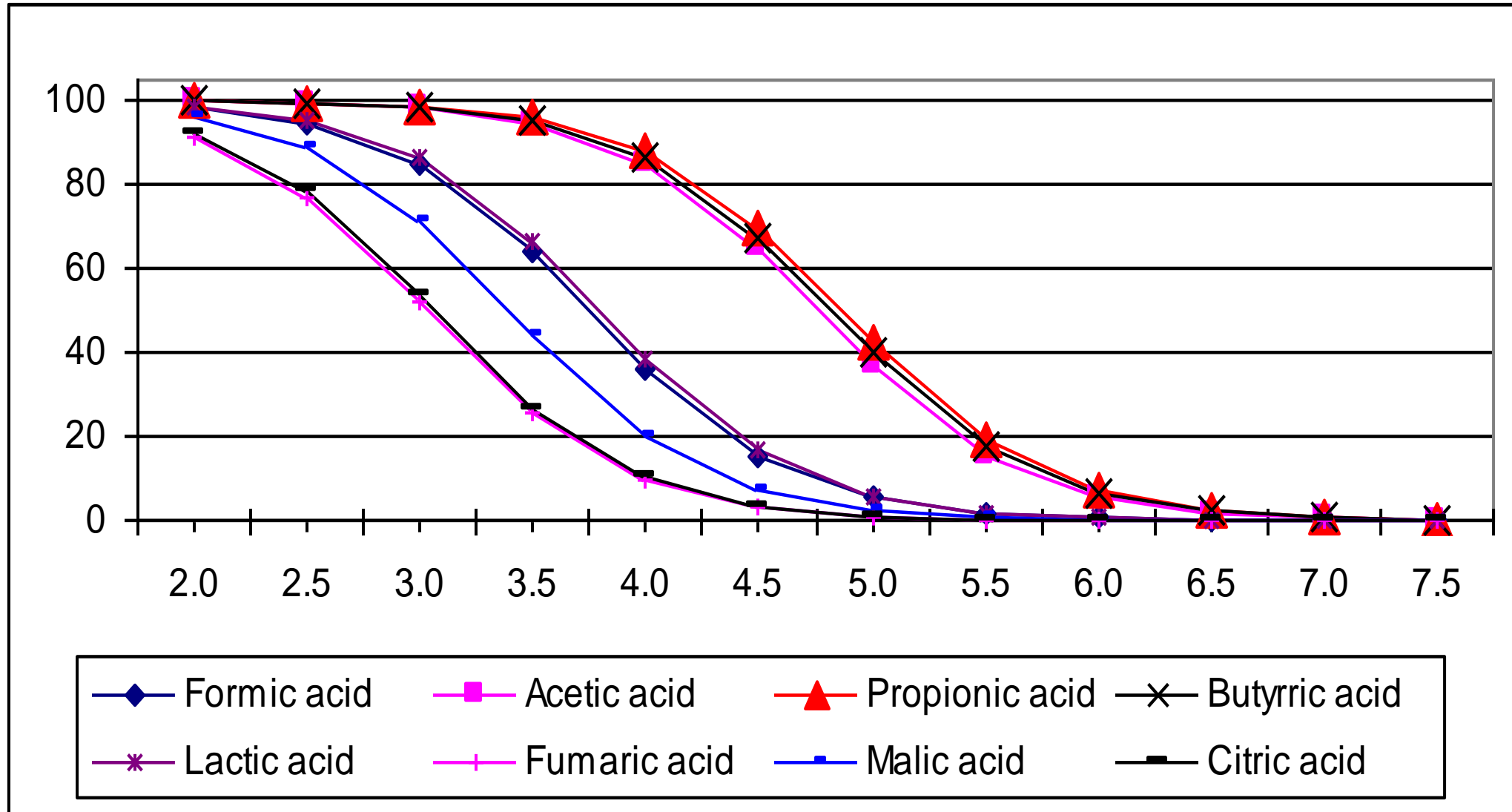
$$\text{pH} = \text{pKa} + \log \left( \frac{[\text{A}^-]}{[\text{HA}]} \right)$$

[https://en.wikipedia.org/wiki/Acid\\_dissociation\\_constant](https://en.wikipedia.org/wiki/Acid_dissociation_constant)

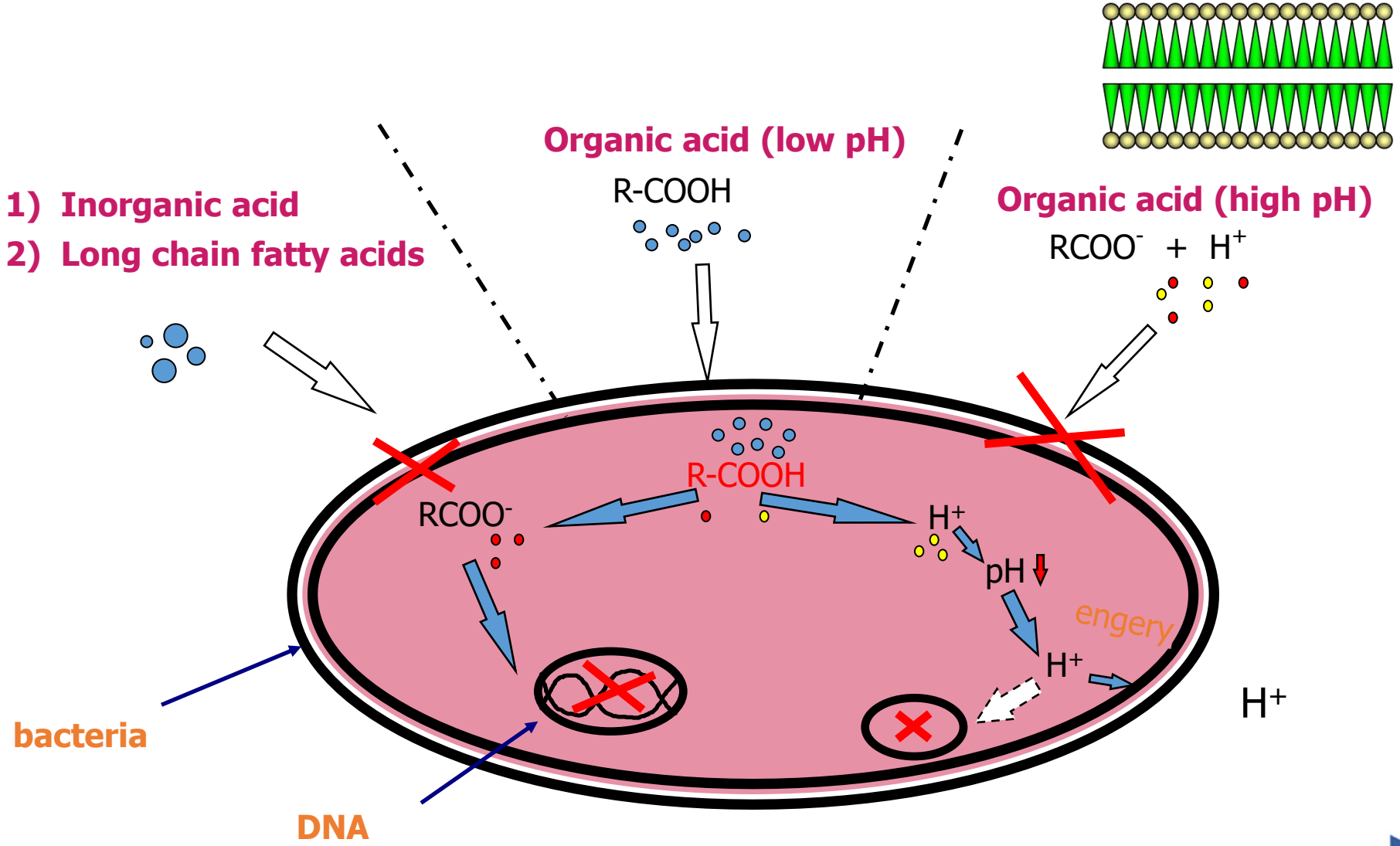
# The pKa of organic acids and HCl

Name	pKa	Name	pKa
Formic acid	3.8	Gallic acid	4.4
Acetic acid	4.8	Malic acid	3.4
Propionic acid	4.9	Fumaric acid	3
Butyric acid	4.8	Lauric acid	5.3
Benzoic acid	4.2	Caproic acid	4.9
Citric acid	3.1	Caprylic acid	4.9
Sorbic acid	4.8	HCl	-7.0

# Relationship between % undissociated acid ([HA]%) and pH



# Mechanisms of pathogen control



(Adapted from R.J. Lambert and M. Stratford, *Journal of Applied Microbiology* 86, 157-164, 1999)

# Experimentally determined values for MICs of undissociated and dissociated organic acids (various authors)

Organism	Acid type	MIC <sub>u</sub> <sup>a</sup>	MIC <sub>d</sub> <sup>b</sup>
<i>E. coli</i> M23	Lactic	8.32	-
<i>Y. enterocolitica</i>	Lactic	5-10	-
<i>E. coli</i>	Propionic	70	800
<i>Staphylococcus aureus</i>	Propionic	19	830
<i>Bacillus cereus</i>	Propionic	17	380
<i>E. coli</i>	Sorbic	1	100
<i>E. coli</i>	Sorbic	1	350
<i>Staphylococcus aureus</i>	Sorbic	0.6	400
<i>Bacillus cereus</i>	Sorbic	1.2	110
<i>Listeria innocua</i>	Lactic (Na lactate)	4.9	1250

<sup>a</sup>MIC<sub>u</sub>, MIC of the undissociated form of acid (micromolar).

<sup>b</sup>MIC<sub>d</sub>, MIC of the dissociated form of acid (micromolar).

Adapted from (Presser)



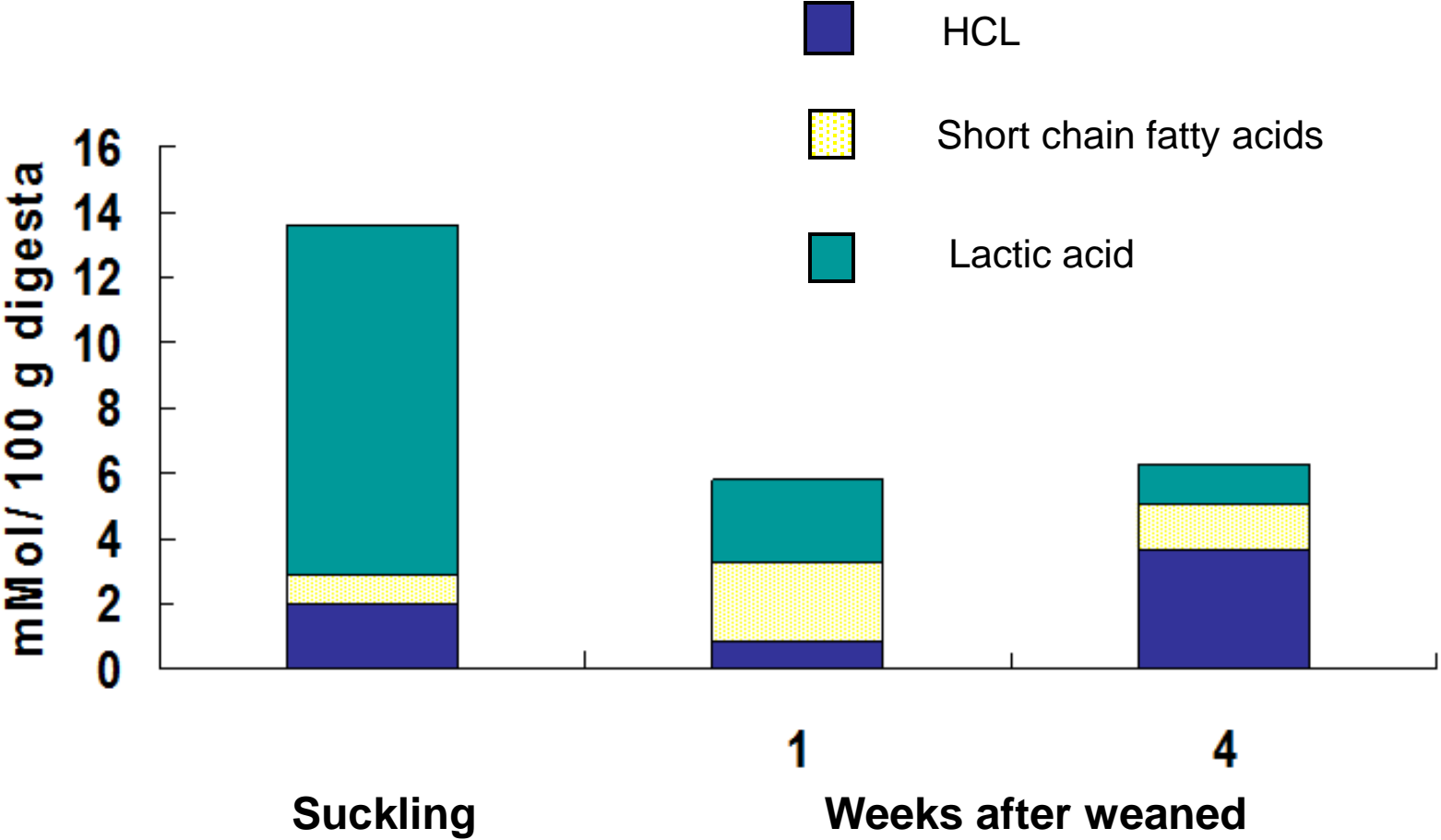
# Why using organic acids in animal feeding?



- 1.- Poor HCL secretion
- 2.- Intestinal pathogen control
- 3.- Buffer capacity of feed



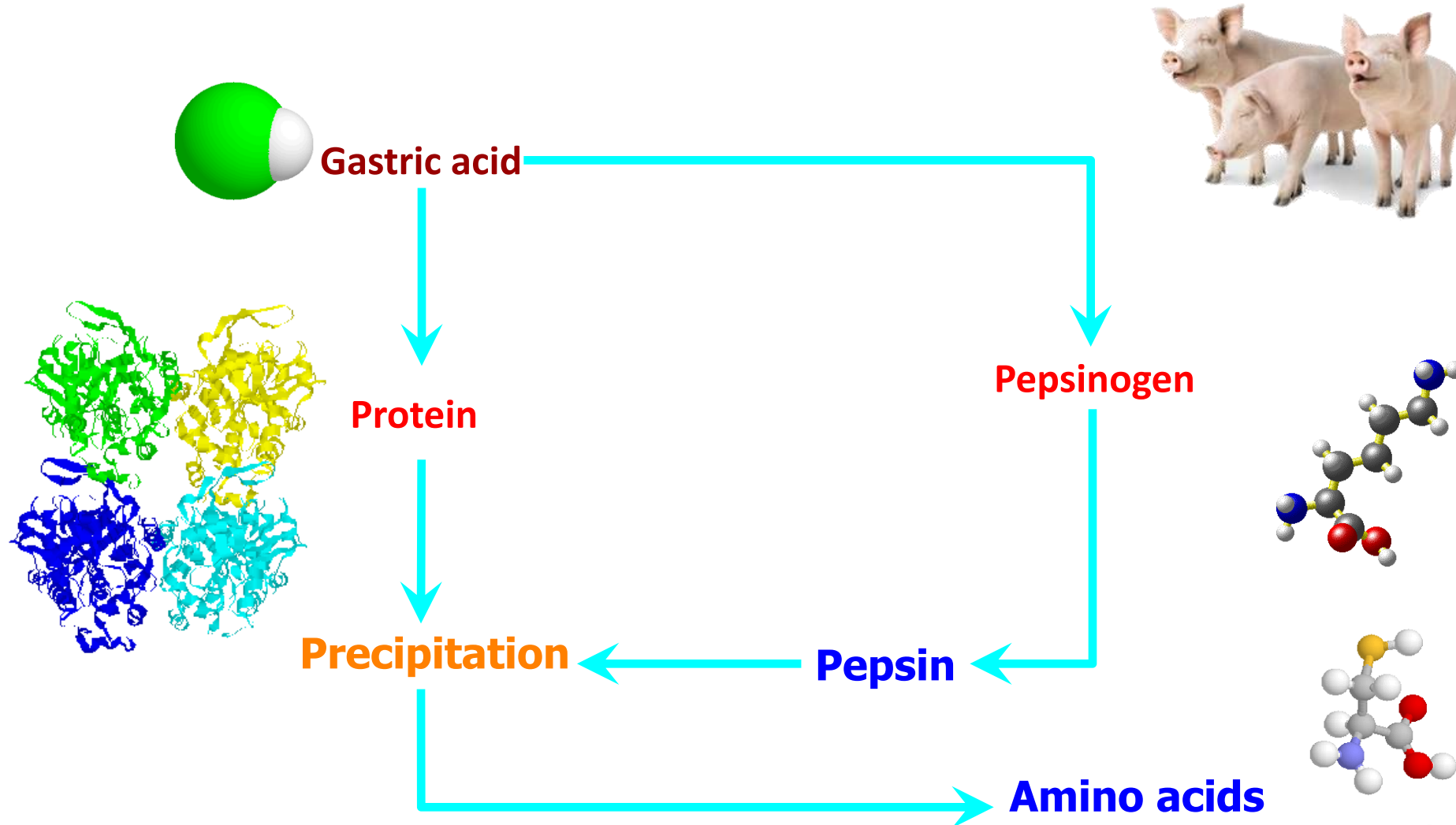
# Decreased HCl secretion in stomach after weaned



*(Weaning at 35 day, Schnabel 1983)*



# Multiple roles of HCl in the stomach



To kill harmful bacteria

To stimulate development of 'good' bacteria (Lactobacilli)



University of Manitoba

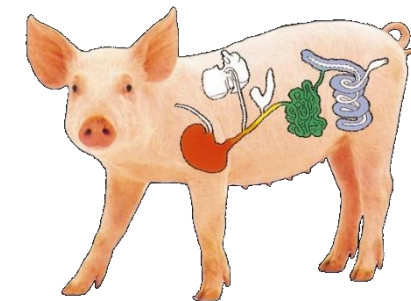
# Acid binding capacity values for different ingredients in piglets

Ingredient	Buffering Capacity, meq/Kg	Ingredient	Buffering Capacity, meq/Kg
Maize	200	L-Lysine	600
Barley	225	DL-Methionine	1000
Wheat Soft	250	L-Threonine	1100
Wheat bran	500	L-Tryptophan	1025
Soybean 44%	1100	Sweet Whey powder	850-1000
Soybean 48%	1100	Betaine	600
Fishmeal 70/72	1800-2200	Dextrose	140-200
Fishmeal Peru origin	1800-2200	Choline	100-150
Vegetable fat	200	Blood plasma	1150-1350
<b>Limestone</b>	<b>18500-22000</b>	<b>ZnO</b>	<b>13000</b>
Calcium formate	9000	<b>Citric acid</b>	<b>-4000</b>
MCP	1800-2000	<b>Fumaric acid</b>	<b>-6400</b>

Definition: Acid Buffering capacity indicates the amount of 1.0 M HCl to lower the pH of feed to a pH =3.

**A target of less than 600 meq/Kg**

(<http://www.wattagnet.com/articles/20464-controlling-dietary-buffering-capacity-in-piglet-feeds>)



# Low calcium levels improve growth in piglets after weaning

Item	Treatments			Pooled SEM	P-Value
	LCa (0.35%)	MCa (0.65%)	HCa (0.95%)		
BW d7, Kg	<b>8.43<sup>a</sup></b>	<b>8.33<sup>ab</sup></b>	<b>8.21<sup>b</sup></b>	0.06	<b>0.041</b>
BW d14, kg	<b>10.72<sup>a</sup></b>	<b>10.6<sup>a</sup></b>	<b>10.16<sup>b</sup></b>	0.08	<b>0.0001</b>
ADG 0–7d, g/d	<b>106.94<sup>a</sup></b>	<b>91.25<sup>ab</sup></b>	<b>74.62<sup>b</sup></b>	8.37	<b>0.044</b>
ADFI 0–7d, g/d	162.51	148.45	149.69	8.05	0.411
G:F 0–7d	0.654 <sup>a</sup>	0.616 <sup>a</sup>	0.481 <sup>b</sup>	0.04	0.014
ADG 0–14d, g/d	<b>219.19<sup>a</sup></b>	<b>207.25<sup>a</sup></b>	<b>176.76<sup>b</sup></b>	5.72	<b>0.0002</b>
ADFI 0–14d, g/d	342.35	323.01	314.64	9.04	0.113

# Different zinc sources may affect the acid buffering capacity of weanling pig diets

	Commercially available protected zinc oxide products			
Acid-binding Capacity	# 1	# 2	# 3	# 4
pH 4	59.43	3785.32	1.91	0.01
pH 3	106.27	3872.62	7.69	0.61
Buffering Capacity				
pH 4	32.92	717.17	0.84	0.01
pH 3	37.82	616.82	2.38	0.15

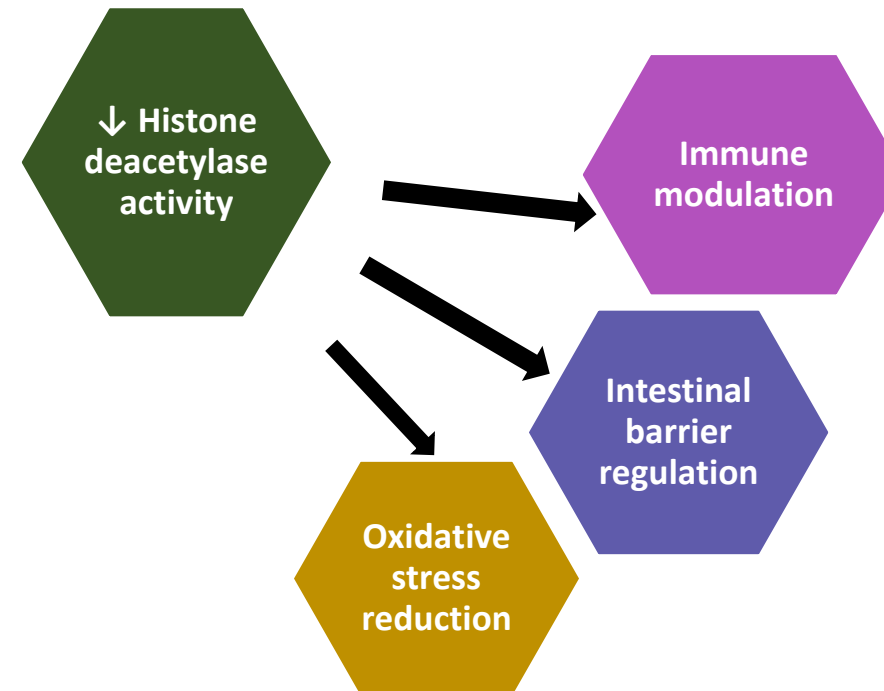
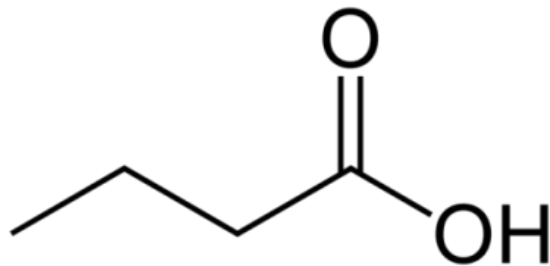
**Acid binding capacity and buffering capacity mean of four zinc products at pH -4 and -3 calculated to 1 mol of the zinc ions.**

*Brown, N. \*, Wang, Y. \*, and Yang, C. Different zinc sources may affect the acid buffering capacity of weanling pig diets. Canadian Hog Journal. Special Summer Edition, For the love of science 2019, P 39-41.*

# Major benefits of butyrate

## Potential alternative to antibiotics (gut health):

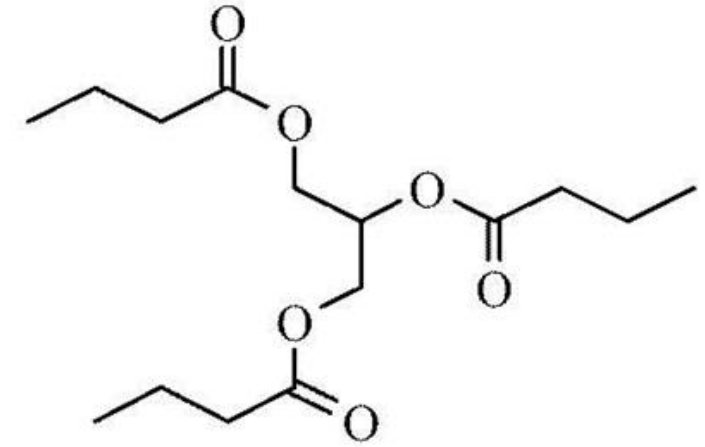
- Anti-inflammatory activity (more energy for growth)
- **Antimicrobials: control of enteric bacterial infection**
- **↑Animal growth performance (mainly on young animals)**
- Histone deacetylase inhibitor



# Limitations and solutions of butyrate in application

## Limitations:

- Offensive odour
- Absorbed to the upper gut



Tri-butyrin

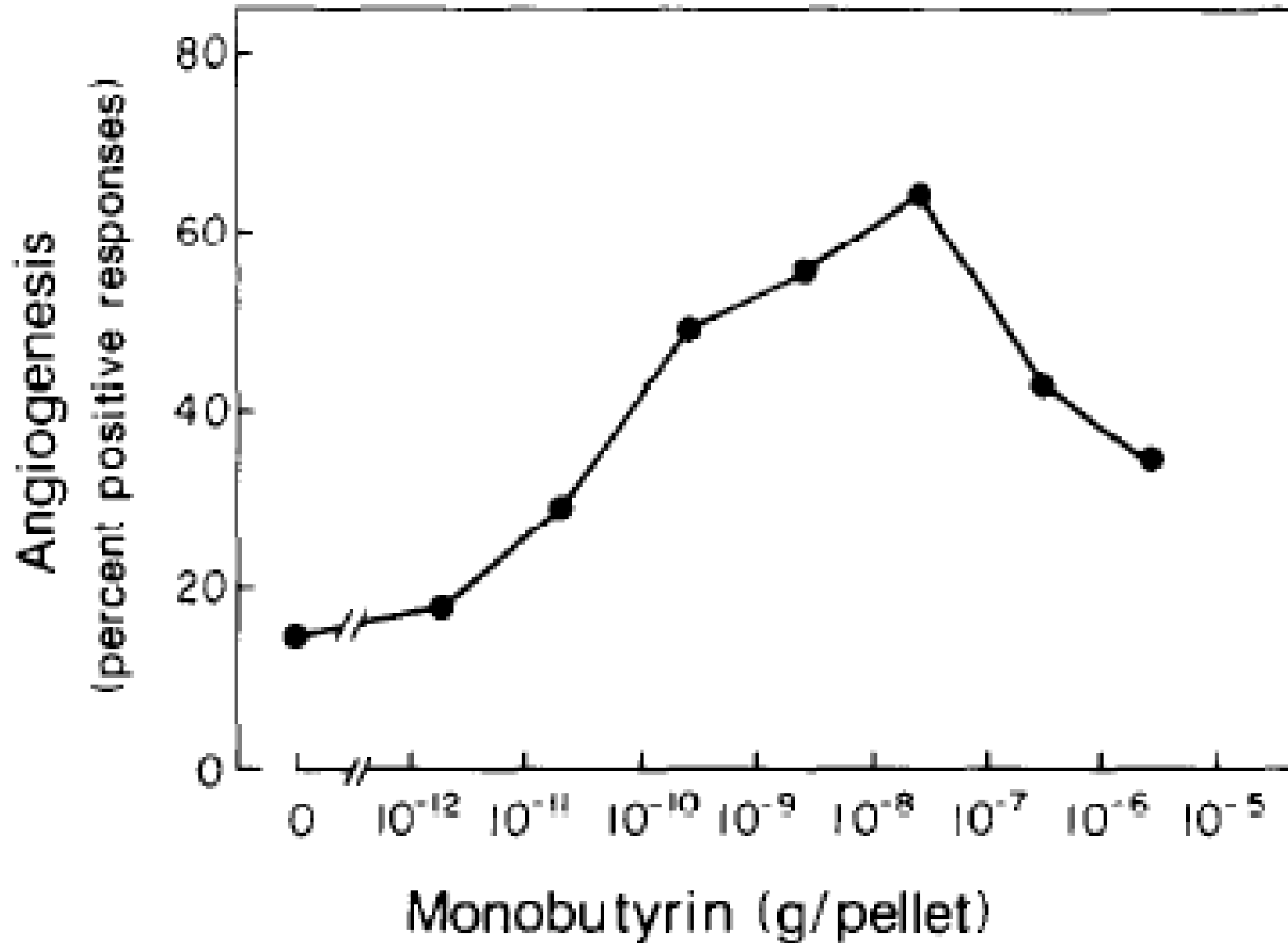
## Solutions:

- Sodium butyrate
- Encapsulated butyrate
- Butyrate glycerides (released by lipase)



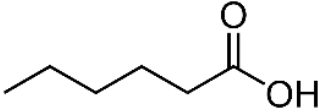
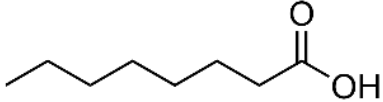
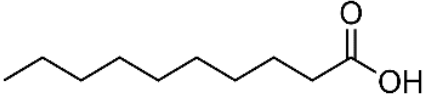
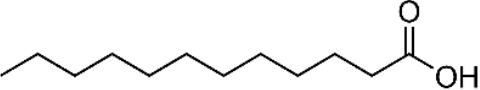


# Angiogenic activity of synthetic and endogenous monobutyryn



*(Dobson et al., 1990; Cell)*

# Molecular structure and physicochemical properties of medium chain fatty acids used in pig feeds

Name	Systemic name	Formula	Skeletal structure	Melting point (° C)	Boiling point (° C)	Density g/mL
Caproic acid	Hexanoic acid	$C_6H_{12}O_2$		-3.4	205.8	0.929
Caprylic acid	Octanoic acid	$C_8H_{16}O_2$		16.7	239	0.910
Capric acid	Decanoic acid	$C_{10}H_{20}O_2$		27-32	268-270	0.893
<b>Lauric acid</b>	<b>Dodecanoic acid</b>	<b><math>C_{12}H_{24}O_2</math></b>		<b>43.8</b>	<b>225-297</b>	<b>1.007</b>

*(Omonijo et al., 2018)*

# Minimal inhibitory concentrations of 15 fatty acids

Compound	Pneumo- cocci	Strep- tococcus group A	Strep- tococcus beta-he- molytic non-A	Coryne- bacteria	<i>Nocardia asteroides</i>	Micro- cocci	<i>Candida</i>	<i>S. aureus</i>	<i>S. epider- midis</i>	Strep- tococcus group D
Caproic . . . . .	NI	NI	NI	NI	NI	NI	NI	NI	NI	NI
Caprylic . . . . .	NI	NI	NI	NI	NI	NI	NI	NI	NI	NI
Capric . . . . .	1.45	1.45	2.9	1.45	1.45	2.9	2.9	2.9	2.9	5.8
Lauric . . . . .	0.062	0.124	0.249	0.124	0.124	0.624	2.49	2.49	2.49	2.49
Myristic . . . . .	0.218	0.547	2.18	0.437	0.547	0.547	4.37	4.37	2.18	4.37
Myristoleic . . . . .	0.110	0.110	0.110	0.055	0.110	0.220	0.552	0.441	0.441	0.441
Palmitic . . . . .	0.48	3.9	3.9	1.9	NI	1.9	NI	NI	3.9	NI
Palmitoleic . . . . .	0.024	0.098	0.049	0.049	0.049	0.049	0.491	0.983	0.491	0.491
Stearic . . . . .	NI	NI	NI	NI	NI	NI	NI	NI	NI	NI
Oleic . . . . .	NI	1.77	NI	NI	NI	NI	NI	NI	NI	NI
Elaidic . . . . .	NI	NI	NI	NI	NI	NI	NI	NI	NI	NI
Linoleic . . . . .	0.044	0.089	0.089	0.044	0.089	0.089	0.455	NI	NI	NI
Linolenic . . . . .	0.179	0.35	0.35	0.179	0.448	0.448	NI	1.79	NI	NI
Linolelaidic . . . . .	NI	NI	NI	NI	NI	NI	NI	NI	NI	NI
Arachidonic . . . . .	NI	NI	NI	NI	NI	NI	NI	NI	NI	NI

<sup>a</sup> Results are given in micromoles per milliliter. NI = not inhibitory at the concentrations tested (1.0 mg/ml or 3 to 6.0  $\mu$ moles/ml).

(Kabara, et al., 1972. *Antimicrobial Agents and Chemotherapy*)

# Minimal inhibitory concentrations of free fatty acids when compared with glycerol forms

Compound	Pneumococci	Streptococcus group A	Streptococcus beta-hemolytic non-A	Corynebacteria	<i>Nocardia asteroides</i>	Micrococci	<i>Candida</i>	<i>S. aureus</i>	<i>S. epidermidis</i>	Streptococcus group D
Capric acid...	1.45	1.45	2.9	1.45	1.45	2.9	2.9	2.9	2.9	5.8
1-Monocaprin.....	0.1	0.2	0.2	0.2	0.5	0.1	1.0	1.0	1.0	2.0
1,3-Dicaprin..	NI	NI	NI	NI	NI	NI	NI	NI	NI	NI
Lauric acid...	0.062	0.124	0.249	0.124	0.124	0.624	2.49	2.49	2.49	2.49
1-Monolaurin.....	0.09	0.045	0.09	0.045	0.09	0.09	0.09	0.09	0.09	NI
1,3-Dilaurin..	NI	NI	NI	NI	NI	NI	NI	NI	NI	NI
Trilaurin.....	NI	NI	NI	NI	NI	NI	NI	NI	NI	NI

<sup>a</sup> Results are given in micromoles per milliliter. NI = no inhibition at the concentrations tested.

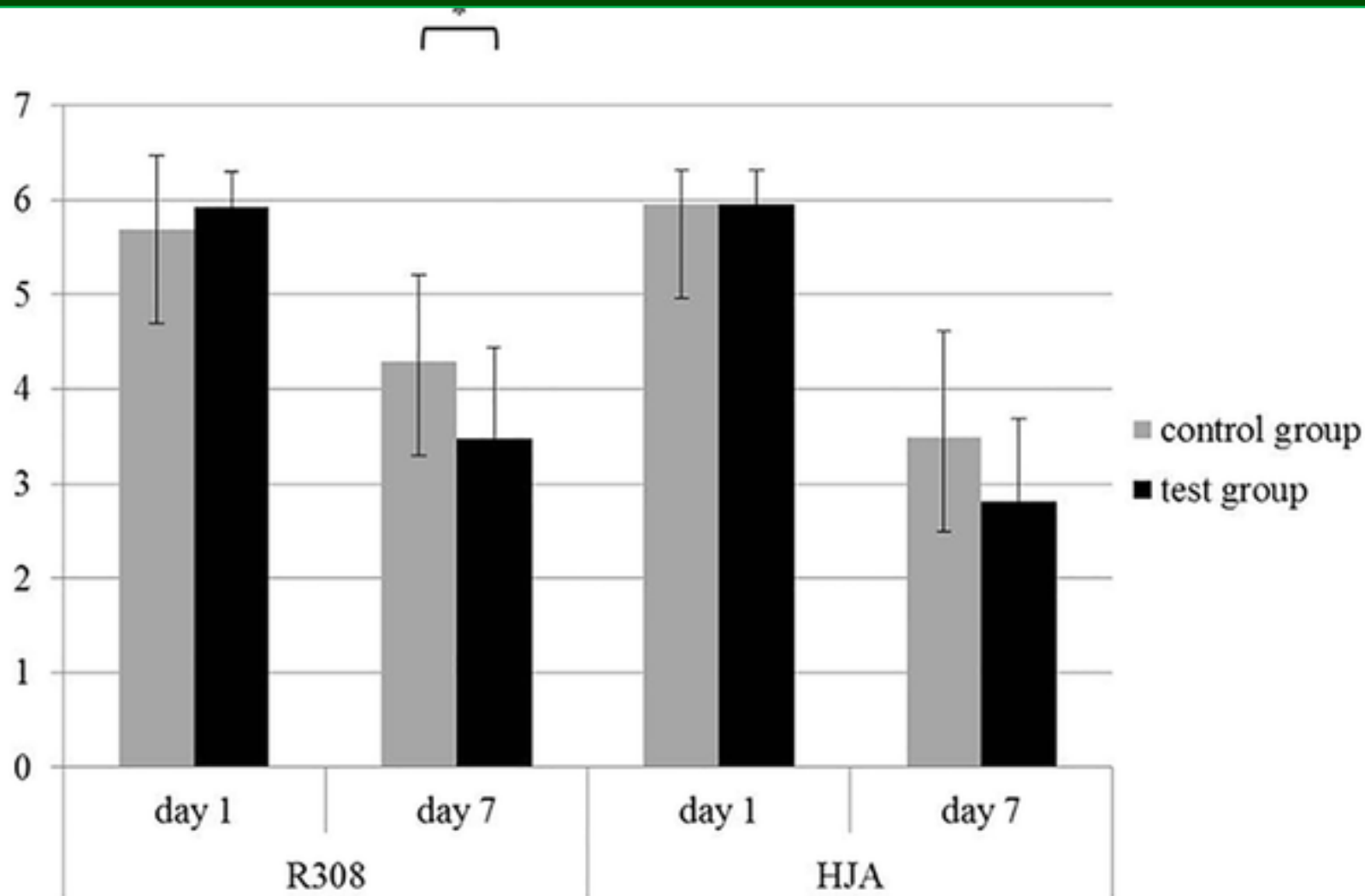
(Kabara, et al., 1972. *Antimicrobial Agents and Chemotherapy*)

# Antibacterial activity of fatty acids against *C. difficile*

	Number of carbon backbone	General name	MIC (mg/ml)
SCFAs	C3	Propionic acid	1.25
	C4	Butyric acid	25
	C4	Isobutyric acid	1.25
	C5	Valeric acid	1.25
	C5	Isovaleric acid	2.5
MCFAs	C6	Hexanoic acid	1.25
	C8	Octanoic acid	2.5
	C10	Capric acid	0.63
	<b>C12</b>	<b>Lauric acid</b>	<b>0.31</b>
LCFAs	C14	Myristic acid	>10
	C16	Palmitic acid	10

(Yang et al., *Front Microbiol.* 2017; 8: 2635)

# Lauric acid reducing *Campylobacter spp.* in broiler meat



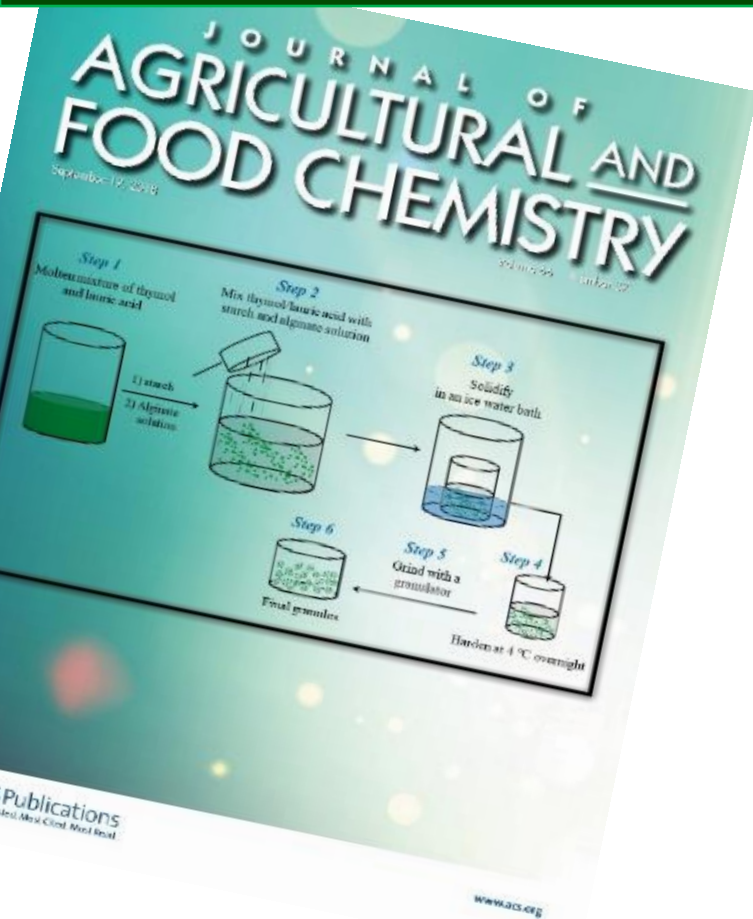
Broilers fed with 5% commercial fat addition (control group) and broilers fed with 5% supplement of fat with high lauric acid content (test group).

# Performance of broiler chickens fed with an experimental diet containing glycerol monolaurate (GML) to replace antibiotic drugs

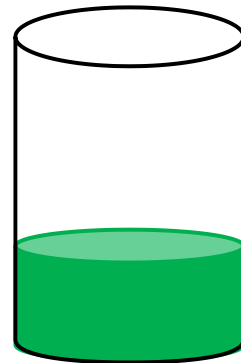
Variables	Days	T0	T100	T200	T300	P value
Weight (g)	1	46.8	48.3	47.7	48.6	0.963
	7	137.9	134.2	135	140.2	0.742
	14	404.4	392	392.2	399.9	0.654
	21	832.9	844.5	800.6	838	0.429
	35	2032.9	2200.6	2153	2204.6	0.175
	42	2598.6b	2756.4ab	2755.6ab	2925.0a	0.001
Feed consumption (g)	42	4196.3	4351.9	4240.5	4422.9	0.098
<b>Weight gain (g)</b>	<b>42</b>	<b>2551.8b</b>	<b>2708.1ab</b>	<b>2707.9ab</b>	<b>2876.5a</b>	<b>0.001*</b>
<b>Daily weight gain (g)</b>	<b>–</b>	<b>61.8b</b>	<b>65.6ab</b>	<b>65.6ab</b>	<b>69.6a</b>	<b>0.001</b>
<b>Feed conversion</b>		<b>1.64a</b>	<b>1.60ab</b>	<b>1.56ab</b>	<b>1.54b</b>	<b>0.036</b>
Mortality (%)	1 to 42	0	0	1.69	5	–

*(Fortuos et al., 2019)*

# Production of thymol and lauric acid microparticle

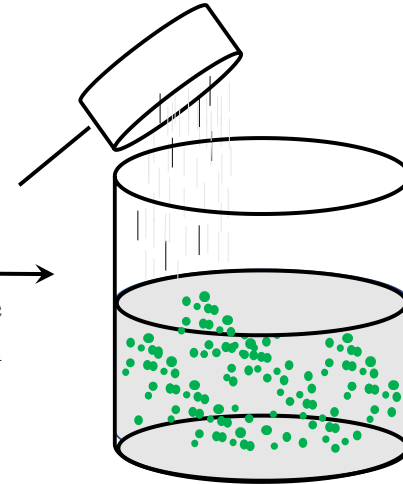


**Step 1**  
Molten mixture of thymol and lauric acid

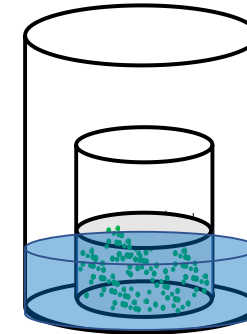


1) starch  
2) Alginate solution

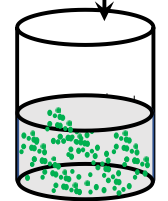
**Step 2**  
Mix thymol/lauric acid with starch and alginate solution



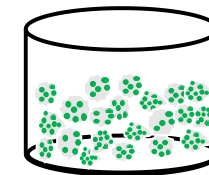
**Step 3**  
Solidify in an ice water bath



**Step 4**  
Harden at 4 °C overnight



**Step 6**



Final granules

**Step 5**  
Grind with a granulator

## Development of Novel Microparticles for Effective Delivery of Thymol and Lauric Acid to Pig Intestinal Tract

Faith A. Omonijo,<sup>†</sup> Seungil Kim,<sup>‡</sup> Tracy Guo,<sup>||</sup> Qi Wang,<sup>||</sup> Joshua Gong,<sup>||</sup> Ludovic Lahaye,<sup>+</sup> Jean-Christophe Bodin,<sup>+</sup> Martin Nyachoti,<sup>†</sup> Song Liu,<sup>‡,§,⊙</sup> and Chengbo Yang<sup>\*,†,⊙</sup>

PCT/CA2019/050599. 2019/05/06



# Pictures of the molten mixture of thymol and FAs (lauric acid, stearic acid and palmitic acid respectively)

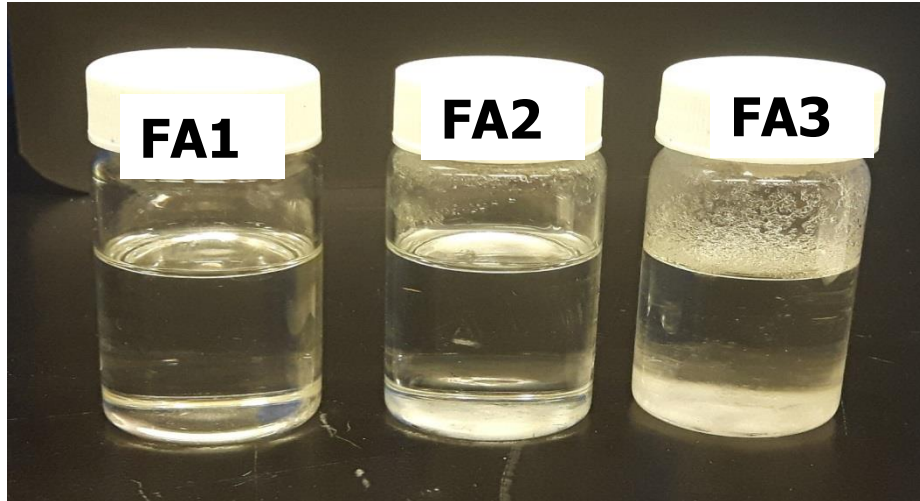


Fig.1. Picture of thymol and FAs mixture set at room temperature at 0 min

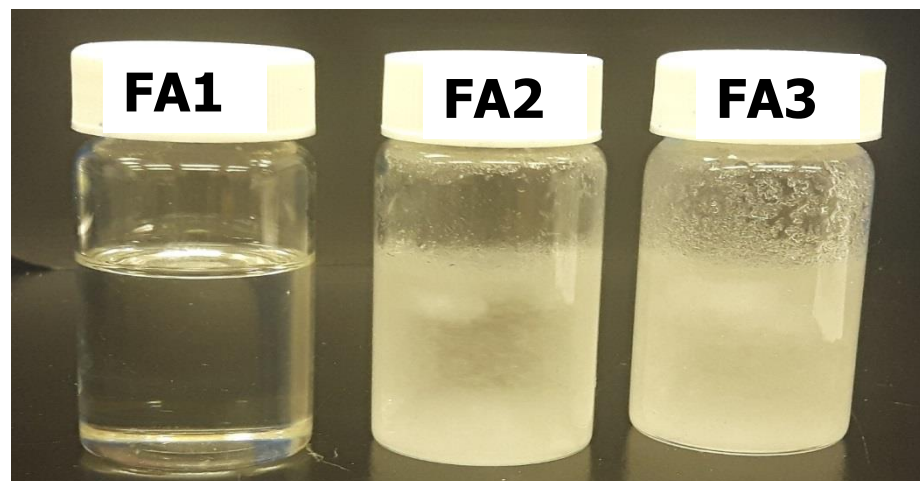


Fig.2. Picture of thymol and FAs mixture set at room temperature at 6 hrs.

## At 0 minute:

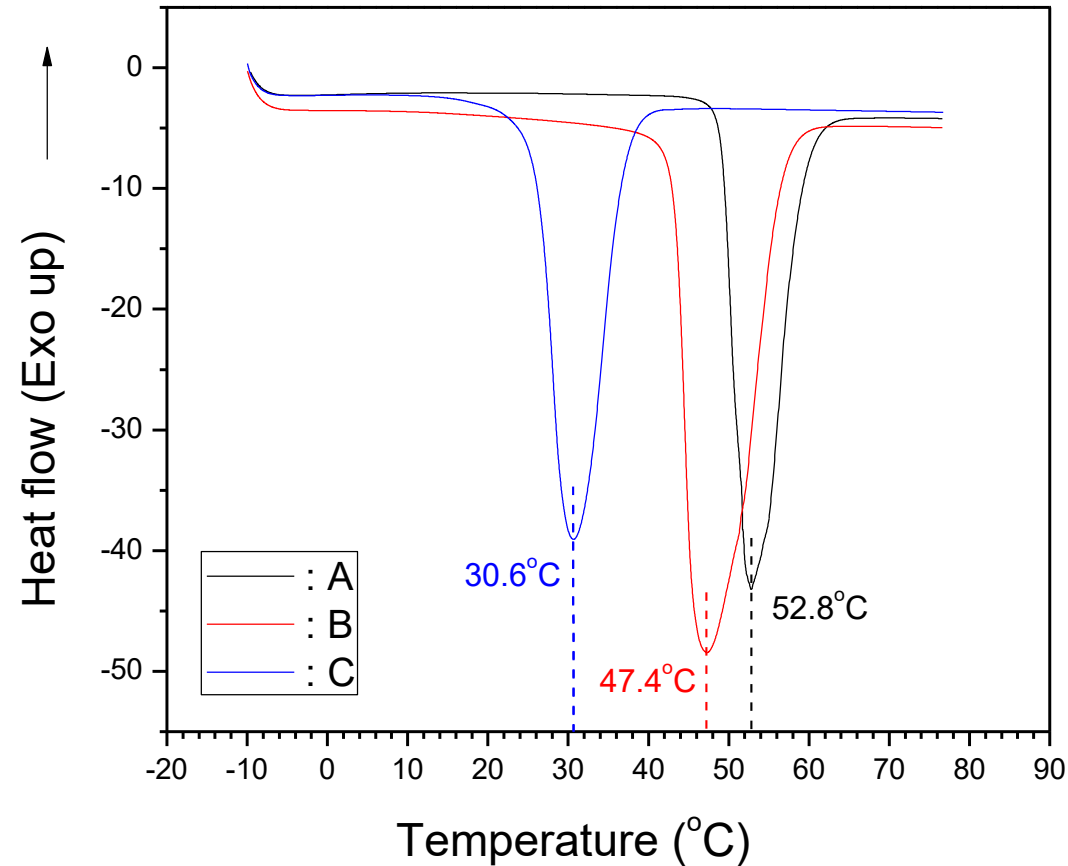
- no phase separation occurred between the mixture.

## After 6 hours:

- Lauric acid and thymol were in liquid state without phase separation
- FA2 and FA3 solidifies.

*(Omonijo et al., 2018)*

Differential scanning calorimetry (DSC) of (A) thymol, (B) lauric acid, and (C) Mixture of thymol and lauric acid (50: 50wt%). 2nd run with heating rate 10 °C/min from -10 °C to 80 °C.

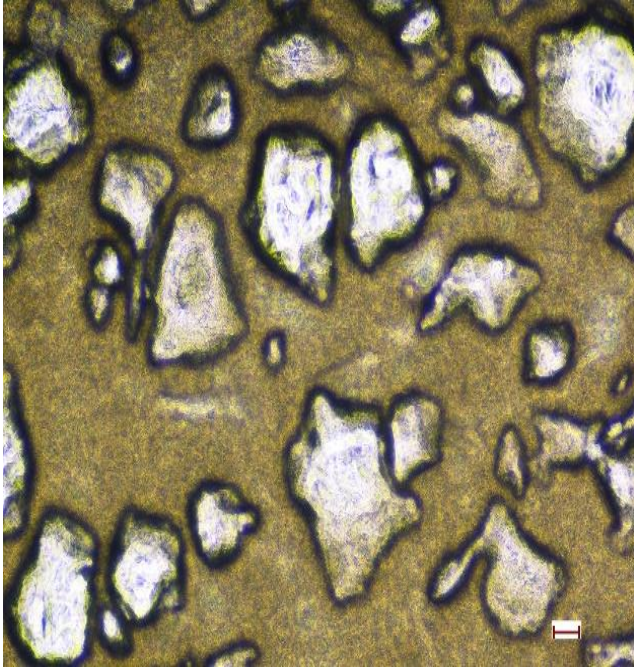


- The melting point of thymol and lauric acid mixture reduced drastically as shown in the graph.
- Explains their eutectic mixture because of their miscibility in liquid state and immiscibility in solid state.

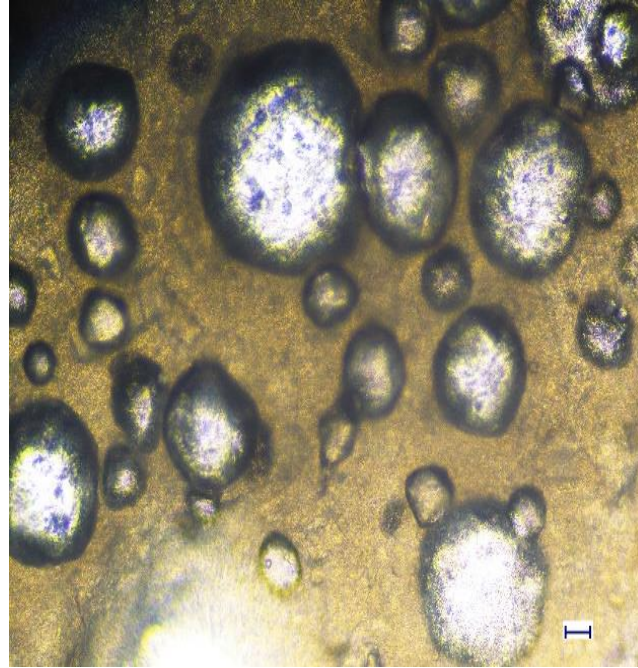
*(Omonijo et al., 2018)*

# Morphology of crystals of thymol (A) and lauric acid (B) and a mixture of thymol and lauric acid (C) after crystallization

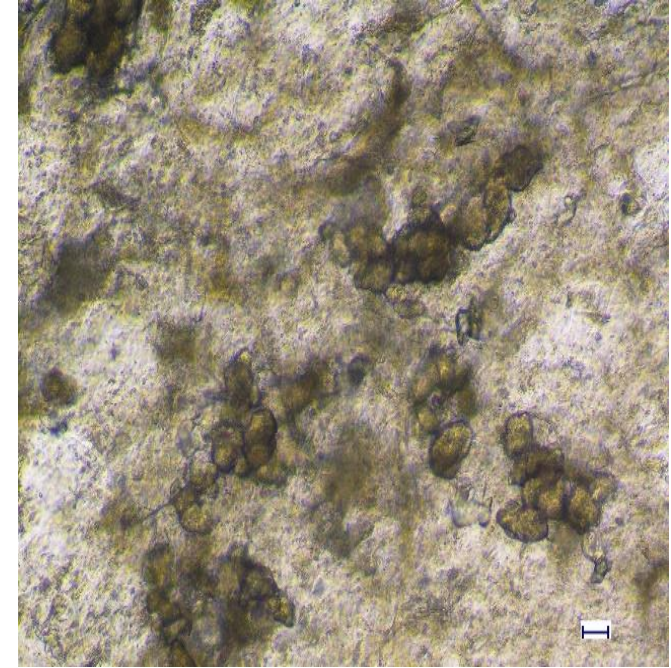
A



B



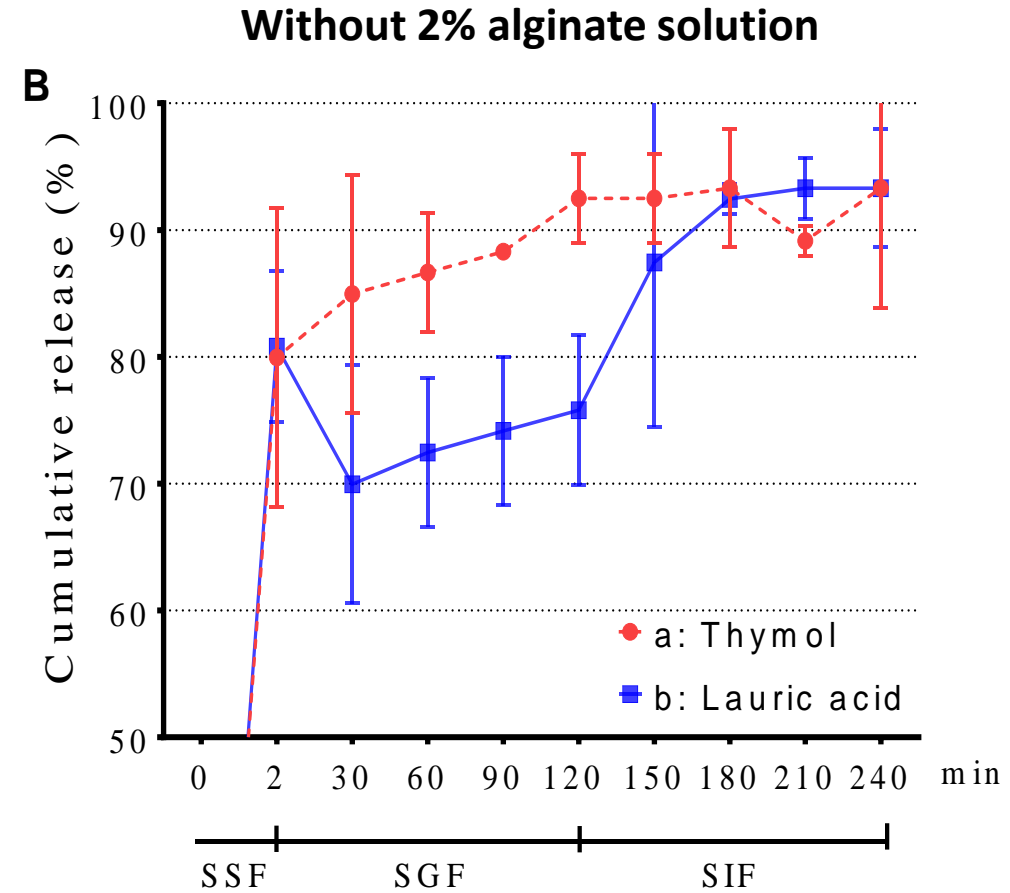
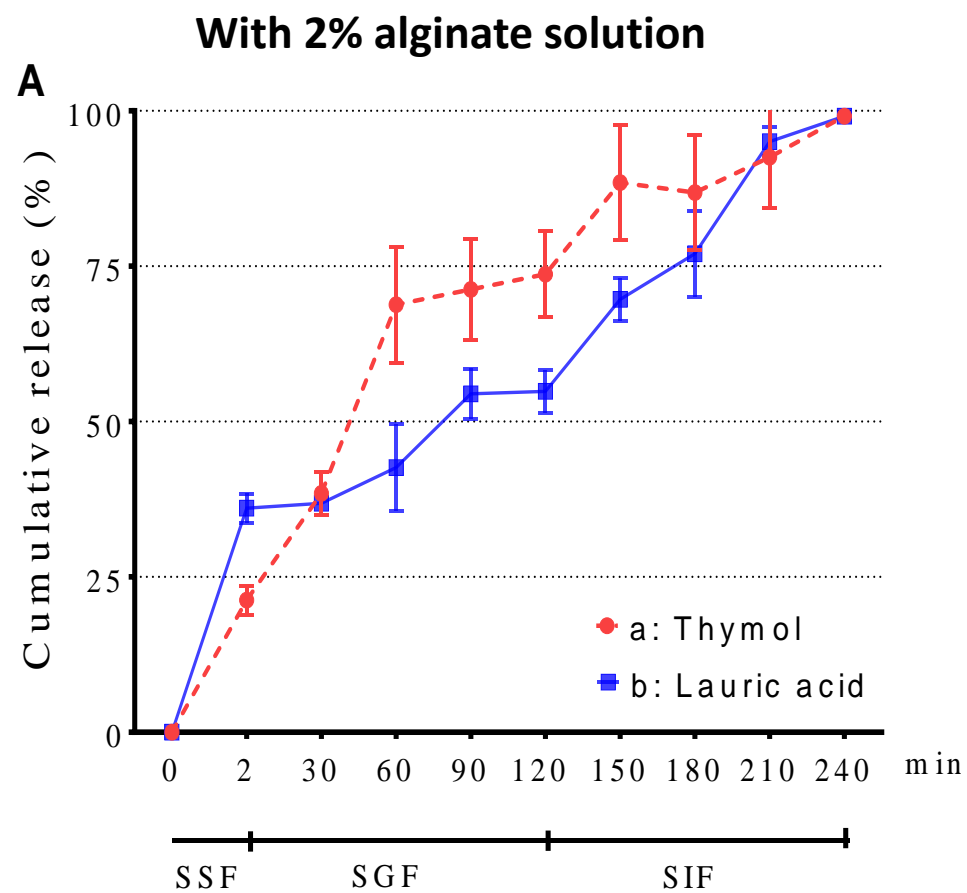
C



- Thymol crystallized in irregular shapes.
- Lauric acid crystallized in round shapes.
- The mixture crystallized into somewhat ovular shaped particles.
- No visible distinctions between the two individual components.

*(Omonijo et al., 2018)*

# *In vitro* release of thymol and lauric acid from microparticles with/without 2% alginate solution



- Microparticles with 2% alginate exhibited slow release compared with microparticles without 2% alginate solution

**SSF:** Simulated salivary fluid  
**SGF:** Simulated Gastric Fluid  
**SIF:** Simulated Intestinal Fluid

*(Omonijo et al., 2018)*

# Conclusions

- **The antimicrobial activity of organic acids is related to environmental pH;**
- **Manipulating dietary acid-binding capacity using organic acids and selecting proper ingredients is considered a possible approach to reduce the use of antibiotics;**
- **Butyrate can improve gut development, gut integrity and immunity;**
- **Protected organic acids or butyrate glycerides can avoid offensive odour;**
- **Monobutyrate glycerides may represent a therapeutic opportunity for stimulating the growth of intestinal tissue through its angiogenic activity in food-producing animals, especially when there are wounds or damages in the intestinal epithelia;**
- **Lauric acid and its derivatives can be used as antibiotic alternatives in livestock production.**

# Acknowledgments

## Fundamental Research



- Discovery Program
- CRD program
- Engage program
- Early Career Researchers (ECR) Supplement



## Current grant support

### Seed grants



- Start-up Funds
- UGRP
- UCRP



Swine Innovation Porc



## Applied Research



# Acknowledgments



# Thank you

